# Multiple Paternity and Breeding System in the Gopher Tortoise, Gopherus polyphemus 

Jamie C. Moon, Earl D. McCoy, Henry R. Mushinsky, and Stephen A. Karl<br>From the Department of Biology, University of South Florida, SCA II0, 4202 East Fowler Avenue, Tampa, FL 33620. Jamie C. Moon is now at the Department of Biology, University of North Florida, 4567 St. John's Bluff Road South, Jacksonville, FL 32224. Stephen A. Karl is now at the Hawai'i Institute of Marine Biology, University of Hawaii, Manoa, PO Box 1346, Kane'ohe, HI 96744.

Address correspondence to S. A. Karl at the address above, or e-mail: skarl@hawaii.edu.


#### Abstract

Little is known about the reproductive behaviors and the actual outcomes of mating attempts in the gopher tortoise (Gopherus polyphemus). We examined the mating system and reproductive behaviors of a population of gopher tortoises in central Florida. Using microsatellite markers, we assigned fathers to the offspring of seven clutches and determined that multiple fathers were present in two of the seven clutches examined. We found that gopher tortoises exhibited a promiscuous mating system with larger males fertilizing the majority of clutches. The advantage of larger males over smaller males in fertilizing females may be a result of larger males winning access to females in aggressive bouts with other males or larger males may be more attractive to females. Clutches produced by larger females tended to be sired by a single male, whereas clutches of smaller females tended to be sired by multiple males.


The gopher tortoise (Gopherus polyphemus) is one of four native North American tortoise species generally occupying dry, sandy habitats of the southern United States and northern Mexico. G. polyphemus ranges in the southeast United States south from the extreme corner of southwestern South Carolina to the tip of the Florida peninsula, west to Louisiana, and north to about the middle of Mississippi, Alabama, and Georgia (Auffenberg and Franz 1982). Wild tortoises generally live for 40-60 years and become reproductively active at $10-20$ years of age (Epperson and Heise 2003; Mushinsky et al. 1994). Although believed to be previously abundant, the gopher tortoise is declining in numbers, mostly due historically to harvesting and recently to extensive habitat loss (Auffenberg and Franz 1982; Diemer 1986). Consequently, the species is of conservation concern and is protected in a variety of ways throughout its range. G. polyphemus internationally is listed in Appendix II of the Convention on International Trade in Endangered Species of Wild Fauna and Flora (Inskipp and Gillett 2003); federally is listed by the U.S. Fish and Wildlife Service as threatened in Louisiana, Mississippi, and west of the Tombigbee and Mobile Rivers in Alabama (U.S. Fish and Wildlife Service 1986); and in the state of Florida is considered a Species of Special Concern by the Florida Fish and Wildlife Conservation Commission (2005).

Habitat loss in Florida is occurring at an alarming rate. Between 1936 and 1987, urban land area increased 500\% in Florida (Kautz 1993), and since 1970, the human population of Florida has increased by more than 3 million persons every decade (Bureau of Economic and Business Research 2000). Also between 1936 and 1987, longleaf pine forest, the prime tortoise habitat, declined in area by $88 \%$ (Kautz 1993) with the remaining forests area now small and fragmented (Noss 1989). Because of these trends, aggressive conservation efforts are needed to ensure the survival of the gopher tortoise.

In general, sound conservation efforts should include as much information as possible on the biology and ecology of a species of interest. In particular, a detailed understanding of the elements of the mating system can be critical to the persistence of a species. How mates are selected determines in large part the fate of genetic variation, and genetic variation, in turn, factors into the presumed long-term evolutionary health of species (Dudash and Fenster 2000; Gilpin and Soulé 1986; Lande and Barrowclough 1987). For turtles in general, monogamous pair-bond formation is uncommon (Pearse and Avise 2001). Parental investment is minimal, and the male's contribution to the offspring is principally the donation of sperm. To the contrary, more promiscuous types of
mating systems, such as polygyny and polyandry, are common in turtles (Crim et al. 2002; Jessop et al. 1999; Valenzuela 2000). In a polygynous system, a single male copulates with multiple females, whereas in a polyandrous system, a single female copulates with multiple males. There exist, therefore, differing expectations for patterns of paternity in clutches produced in polygynous versus polyandrous systems; multiple paternity should be the norm in polyandrous systems but not necessarily so in polygynous systems. Although female turtles apparently do not gain direct mating benefits, such as nuptial gifts or paternal care of offspring, possible indirect benefits to mating with more than one male would include gaining "good genes" (Kempenaers et al. 1992; Otter and Radcliffe 1996; Watson 1998), avoidance of genetic incompatibility (Zeh JA and Zeh DW 1996), fertility insurance (Orsetti and Rutowski 2003), and increased genetic diversity of offspring (Byrne and Roberts 2000; Madsen et al. 1992).

Many turtle species are known to store sperm for long periods of time. For example, a captive female box turtle continued to produce viable offspring 4 years after being isolated from males and thus a source of sperm (Ewing 1943). The desert tortoise (Gopherus agassiziz) has been shown to store sperm for at least 2 years (Palmer et al. 1998; Pearse and Avise 2001). Because of sperm storage, females could engage in a form of temporal polyandry. Multiple paternity in gopher tortoises, therefore, could result from inter- as well as intraannual copulations. Gopher tortoises exhibit mating behaviors from March to September; egg laying, however, is only known to occur in May and June (Butler and Hull 1996; Douglass 1976; Iverson 1980). It seems likely then that sperm from late summer or fall mating is stored and used to fertilize eggs the next spring. Therefore, even females mating with only a single male within a season, but not between seasons, would be effectively polyandrous. Clearly, sperm storage could lead to a higher probability or degree of multiple paternity even in a strictly monogamous species. Thus, multiple paternity in gopher tortoises may be an example of temporal as well as single-season polyandry, the combined effect of which can be estimated indirectly by assessing the degree of multiple paternity of natural clutches.

Very little is known about the mating system of gopher tortoises. Male dominance hierarchies as related to mating have been documented; larger males most often are the victors in aggressive male-male interactions (Douglass 1976; McRae et al. 1981). Douglass (1976) hypothesized that dominant males might also maintain a loose harem (but see Boglioli et al. 2003). Given the general low density of females and unfavorable energetics, McRae et al. (1981) suggested that it would be unlikely for a male to defend several courted females, and the results of a study by Boglioli et al. (2003) supported scramble competition polygyny. Instances of male courtship behaviors (e.g., head bobbing) have been observed and are commonly reported in wild tortoise populations, although subsequent copulation rarely has been observed except in captive settings (Auffenburg 1966; Douglass 1976; Wright 1982). Male-male confrontations, in which the participants attempt to overturn one another, also are observed during courting season (Hailman 1991; personal observa-
tion). The importance of these behaviors, however, is largely unknown. Field observation can identify dominant males on an incident-by-incident basis, but rarely can they reveal whether dominance leads to greater reproductive success. For this, an assessment of the actual paternity of the offspring in a clutch is necessary. Molecular genetic techniques able to identify single individuals are providing answers to a variety of questions concerning mating systems, such as paternity (Roques et al. 2004), maternity (Jones et al. 1999), and mating behaviors (Strausberger and Ashley 2003). Microsatellite loci are by far the most frequently used genetic tool in the investigation of specifics of mating systems. These loci mutate quickly and, therefore, are highly polymorphic (Dallas 1992) making them especially useful in identifying individuals (Queller et al. 1993). The probability of identifying or excluding from consideration specific individuals often is extremely large with multilocus microsatellite genotypes. Given this, microsatellite markers are an ideal genetic tool for assaying multiple paternity.

Here, we report the results of a microsatellite locus genotyping study examining the mating system of a population of gopher tortoises in central Florida. In particular, we addressed the following questions: Does multiple paternity occur in gopher tortoise clutches, and, if so, is the fertilization of a single clutch evenly divided among the males? Do some males fertilize more clutches than others, and, if so, is fertilization success related to male size?

## Methods

## Sample Collection

Our field site was the University of South Florida Ecological Research Area (ERA), a 200-ha reserve located in Hillsborough County in west central Florida $\left(28.05^{\circ} \mathrm{N}, 82.20^{\circ} \mathrm{W}\right)$. Approximately 20 ha of sandhill habitat within the ERA have been exposed to controlled burning since 1976 (Mushinsky 1992). This area is separated into 11 plots that are burned on 1 -, 2 -, 5 -, or 7 -year cycles or left as unburned controls (Mushinsky 1985). Tortoises are distributed through the upland habitat of the ERA but are most commonly found in the plots subjected to control burns.

A population of about 280 tortoises occupies the ERA (Mushinsky et al. 1994). All plots were trapped for tortoises during the course of this study from April to August of 2001 and again in 2002, and all active and inactive burrows (based on Mushinsky and McCoy 1994) in each plot were located and marked. The width of each burrow was measured at a depth of 500 mm and used as an estimate of the carapace length (CL) of the resident tortoise (Wilson et al. 1991). Burrows greater in width than the minimum CL of a sexually mature individual of either sex (i.e., males at 177 mm ; Diemer and Moore 1994; Mushinsky et al. 1994) were pit trapped by burying at the burrow entrance a $9.5-\mathrm{L}$ bucket camouflaged with brown fabric and sand with the opening level with the surface of the ground. While set, the traps were checked every 2 h during daytime. Individuals also were captured by hand when encountered outside their burrows.

Sex and stage of maturity in gopher tortoises can be inferred readily by the size of the carapace. Tortoises larger than $240-\mathrm{mm}$ CL display sex-specific degrees of plastral concavity (PC). The PC of females is less, and that of males greater, than 6 mm deep (Mushinsky et al. 1994). Females in the ERA are reproductive at $\sim 240-\mathrm{mm}$ CL (Mushinsky et al. 1994) and males at CL from 177 to 230 mm (Diemer and Moore 1994).

Mass and CL were assessed for all sexually mature and subadult tortoises captured (Mushinsky et al. 1994). Differences between various groupings of individuals were tested using Student's $t$ tests. The CL and mass of the mothers and the fathers assigned to the clutch based on the genetic data for single- and multiple-sire clutches, as well as clutch characteristics, also were compared using Student's $t$ tests. A blood sample was obtained from the brachial sinus with a heparinized 21 -gauge needle and a $10-\mathrm{cc}$ syringe. After collection, blood was stored in a lysis buffer ( 10 mM Tris$\mathrm{HCl} \mathrm{pH} 8.0,100 \mathrm{mM} \mathrm{NaCl}, 50 \mathrm{mM}$ ethylenediaminetetraacetic acid, $1 \% \mathrm{w} / \mathrm{v}$ polyvinylpyrrolidone, and $0.2 \% \mathrm{v} / \mathrm{v}$ 2-mercaptoethanol) until DNA extraction. Unless already identified, all individuals were numbered and uniquely marked by notching the marginal scutes, and all individuals were released at the site of capture. The overall goal was to genotype all sexually mature males in the population (i.e., the paternal pool) and genotype and capture all sexually mature females.

To determine if they were carrying shelled eggs, sexually mature females were taken to the laboratory and radiographed with one pulse from The Inspector X-ray Source, Model 200 (Golden Engineering, Centerville, Indiana), with a source output of 3 mR at 1 ft per pulse. Females with completely shelled eggs were given an injection of 1.5 units per $100-\mathrm{g}$ body mass of $3 \%$ oxytocin to stimulate oviposition (Ewert and Legler 1978; Iverson J, personal communication). After oviposition, blood was drawn and stored for DNA extractions, and mothers were returned to the site of capture.

Eggs were incubated in the laboratory in nests of vermiculite with a $1: 1 \mathrm{w} / \mathrm{v}$ ratio of vermiculite to water (Burke et al. 1996) and the temperature maintained at $\sim 30^{\circ} \mathrm{C}$ (Burke et al. 1996; Demuth 2001). Eggs were inspected daily during incubation and hatchlings removed when discovered. Eggs that did not hatch after 120 days were removed and opened to remove the embryo, if present, and frozen for later DNA extraction. Live hatchlings were maintained in aquaria with a 12:12 light:dark cycle and fed a diet of vegetables supplemented with vitamin and mineral powders. Blood samples were obtained from hatchlings by cardiocentesis with a heparinized 26-gauge needle and a 1-cc syringe (Jacobson E, personal communication). Hatchlings were given individual markings in the same manner as adults and released at the burrow where the mother was captured.

## Genotype Analysis

Total cell DNA was extracted following a standard phenol/ chloroform protocol and ethanol precipitated (Herrmann and Frischauf 1987). All individuals were genotyped at nine microsatellite loci (GP15, GP19, GP26, GP30, GP55,

GP61, GP81, GP96, and GP102) previously characterized by Schwartz et al. (2003). Microsatellite polymerase chain reaction amplification reactions were run with one of two multiplexing primer mixes with reaction mixes and cycling parameters following Schwartz et al. (2003). After amplification, samples were run on an ABI Prism 377 automated DNA sequencer (Perkin Elmer, Applied Biosystems, Inc, Foster City, CA) at Iowa State University. The program, GENESCAN (Perkin Elmer, Applied Biosystems, Inc.), was used to identify and quantify microsatellite peaks, and electropherograms of all individuals at all loci were scored once each by three persons to ensure accuracy in reading and recording of results.

Maternal contribution of each individual in the clutch was determined by comparing the offspring's and the mother's genotypes. Paternal alleles were inferred by removing known maternal allelic contributions from the offspring genotypes, when possible. If among all hatchlings, more than two paternal alleles were found at any locus, the clutch was classified as having multiple sires. For multiply sired clutches, paternity exclusion was done on an individual hatchling basis. In any case, candidate males whose genotype mismatched the paternal contribution of the hatchlings at one or more loci were excluded as potential sires. Individual and combined exclusion probabilities for all loci were calculated with the program CERVUS, version 2.0 (Marshall et al. 1998). When more than one male could not be excluded as the father of a hatchling, we performed a probability analysis on all nonexcluded males. CERVUS uses a $\log$ (base e) likelihood algorithm to calculate the likelihood ratio (LOD) of a candidate male being the true parent compared to an arbitrary male. LOD scores are calculated for all possible sires (i.e., nonexcluded males), and the difference between the two most likely candidates $(\Delta)$ is calculated and provides an indication of the reliability of the assignment. By simulation, a critical $\Delta$ score is calculated at either relaxed $(80 \%)$ or strict $(95 \%)$ confidence level and corresponds to an estimated frequency of false positives of $20 \%$ and $5 \%$, respectively. In multiple sire clutches, it is possible that all males except one are excluded as potential fathers for some, but not all, of the hatchlings. When the unequivocally assigned father was one of the nonexcluded fathers for a clutch mate with more than one potential sire, he was assigned as the father regardless of the $\Delta$ value.

Allele frequencies were estimated only from wild-caught individuals (i.e., not including hatchlings). The probability of two unrelated individuals sharing the same genotype at all loci (probability of identity) was calculated as in Hanotte et al. (1991). The probability of exclusion $(D)$ is the overall probability of detecting multiple paternity and was calculated as in Westneat et al. (1987). This calculation utilizes the frequency of alleles and all possible mating arrangements to compute the probability of detecting when a female has mated with another male besides the candidate father, given the loci used.

## Results

We captured 15 adult females representing approximately $31 \%$ of the 49 adult females observed in the ERA between 1988 and 1993. Seven of the 15 were gravid and produced

Table I. Characteristics of mother, clutch, and father for single-sired and multiple-sired clutches

|  | Single sired |  |  |  |  | Multiple sired |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | Clutch 18 | Clutch 201 | Clutch 253 | Clutch 378 | Clutch 529 | Clutch 446 | Clutch 523 |
| Mother ID | 18 | 201 | 253 | 378 | 529 | 446 | 523 |
| Mother CL (mm) | 300 | 314 | 306 | 291 | 317 | 276 | 289 |
| Mother mass (g) | 4400 | 4850 | 4100 | 4300 | 4450 | 3100 | 3100 |
| Father ID | 43 | 343 | 441 | 519 | 346 | $43^{a} \quad 533$ | $180^{a} \quad 256$ |
| Father CL (mm) | 290 | 278 | 250 | 264 | 270 | 290263 | 260276 |
| Father mass (g) | 3800 | 4000 | 2800 | 3500 | 3400 | 38002800 | 28503500 |
| Total number of eggs | 9 | 12 | 9 | 7 | 9 | 6 | 10 |
| Hatching success (\%) | 100.0 | 91.6 | 33.3 | 57.2 | 55.0 | 33.3 | 0.0 |
| Number of undeveloped eggs | 0 | 1 | 0 | 2 | 0 | 1 | 3 |
| Number of eggs genotyped | $8^{b}$ | 11 | 9 | $4^{b}$ | 9 | 5 | 7 |
| $\%$ Fertilized by $1^{\circ}$ male | 100.0 | 100.0 | 100.0 | 100.0 | 100.0 | 80.0 | 57.1 |

${ }^{a}$ This male sired the largest number of the clutch mates and is designated as the primary male.
${ }^{b}$ A blood sample could not be obtained from one member of this clutch.
clutches with the number of offspring per clutch ranging from 4 to 11 with a mean of $7.57 \pm 2.44$ (Table 1). A total of 90 individuals were genotyped, including 5 subadults, 15 adult females, 17 adult males, and 53 offspring. The nine microsatellite loci had two to five alleles each, with a mean of $3.44 \pm 1.01$ alleles per locus and observed per-locus heterozygosity ranged from 0.053 to 0.658 (Table 2 ). The average probability of two unrelated tortoises sharing the same genotype at all nine loci (probability of identity) was $3.84 \times$ $10^{-4}$ (Table 2). The probability of detecting multiple paternity $(D)$ with a single locus ranged from 0.026 (at GP96) to 0.410 (GP81; Table 2) and was 0.876 when all nine loci were used. The cutoff $\Delta$ value estimates for the relaxed and strict confidence limits were $\Delta_{80 \%}=1.44$ and $\Delta_{95 \%}=3.10$, respectively.

## Paternal Assignment

In most cases, the maternal contribution could be unambiguously determined by inspection. In cases where the mother and offspring were both heterozygous for the same alleles, determining unambiguously which allele was the maternal allele was impossible. Males who possessed at least one copy of

Table 2. The number of alleles, observed $\left(H_{\mathrm{O}}\right)$ and expected $\left(H_{\mathrm{E}}\right)$ heterozygosity, probability of identity, and probability of exclusion for nine microsatellites in Gopherus polyphemus

| Locus | Number <br> of alleles | $H_{\mathrm{O}}$ | $\mathrm{H}_{\mathrm{E}}$ | Probability <br> of identity | Probability <br> of exclusion |
| :--- | :--- | :--- | :--- | :--- | :--- |
| GP15 | 5 | 0.605 | 0.611 | 0.210 | 0.359 |
| GP19 | 3 | 0.184 | 0.295 | 0.532 | 0.139 |
| GP26 | 4 | 0.289 | 0.305 | 0.506 | 0.163 |
| GP30 | 3 | 0.421 | 0.500 | 0.366 | 0.199 |
| GP55 | 2 | 0.132 | 0.125 | 0.773 | 0.058 |
| GP61 | 3 | 0.184 | 0.172 | 0.701 | 0.082 |
| GP81 | 5 | 0.658 | 0.662 | 0.168 | 0.410 |
| GP96 | 3 | 0.053 | 0.052 | 0.901 | 0.026 |
| GP102 | 3 | 0.632 | 0.613 | 0.227 | 0.326 |
| Overall | $3.44 \pm 1.01$ | 0.351 | 0.371 | $3.84 \times 10^{-4}$ | 0.876 |

either allele, therefore, could not be excluded from the analysis. When a father was assigned to a clutch or an individual, the probability of another unrelated tortoise having the same genotype as the father was extremely low, ranging from $5.65 \times 10^{-13}$ to $1.37 \times 10^{-6}$ (Table 3).

The genetic contribution of the male also was often unambiguously identified. In four of the seven clutches, all candidate males but one were excluded because, at one or more of the loci, they could not have contributed the paternal allele to the offspring (Table 3). For one hatchling in a fifth clutch (378), all but one male (519) were excluded as potential sires. For each of the remaining three clutch mates, however, three candidate males could not be excluded based on genotype alone. Results from an analysis with CERVUS indicate that male 519 was the most likely father to all three with at least $80 \%$ confidence ( $2.52 \leq \Delta \leq 3.20$ ), so we assigned this male as the sole father of the clutch. In the remaining two clutches, three or more paternal alleles were found at one or more loci, indicating multiple sires for these clutches. For one hatchling in one of these clutches (446), all males except 533 could be excluded as a sire. For the remaining clutch mates, however, male 43 was either the only nonexcluded male (two individuals) or the most likely sire in the likelihood analysis (two individuals) albeit with small $\Delta$ values ( $0.056 \leq \Delta \leq$ 0.064 ). Male 43 also was the sole putative sire for one of the single-sire clutches (Table 3). In the other multiple-sire clutch (523), males 180 and 265 were each either the only nonexcluded male or one of several nonexcluded fathers (Table 3). For one individual in this clutch, all the males included in this survey were excluded based on genotype alone because this hatchling had one allele at each of two loci not seen in any male surveyed.

## Mating System

Reproductive output and hatching success were variable among females. The number of eggs in the clutch did not vary significantly between single- and multiple-sire clutches $\left(t_{\mathrm{df}=5}=0.703, P=.513\right.$; Table 1$)$. The number of undeveloped eggs also did not vary between single- and multiple-sire

Table 3. Results of paternity assignment and probability of identity for the seven clutches and females surveyed in this study

| Clutch number | Number of offspring | Number nonexcluded | Assigned male | $\Delta^{a}$ | Probability of identity |
| :---: | :---: | :---: | :---: | :---: | :---: |
| Putative single sire |  |  |  |  |  |
| 18 | 8 | 1 | 43 | - | $1.14 \times 10^{-10}$ |
| 201 | 11 | 1 | 343 | - | $4.11 \times 10^{-13}$ |
| 253 | 9 | 1 | 441 | - | $5.65 \times 10^{-13}$ |
| 529 | 9 | 1 | 346 | - | $2.82 \times 10^{-8}$ |
| 378 | 1 | 1 | 519 | - | $1.37 \times 10^{-6}$ |
|  | 3 | 3 | 519 | 2.52-3.20 | - |
| Putative multiple sire |  |  |  |  |  |
| 446 | 1 | 1 | 533 | - | $7.07 \times 10^{-12}$ |
|  | 2 | 1 | 43 | - | $1.14 \times 10^{-10}$ |
|  | 2 | $4^{b}$ | 43 | 0.06 | - |
| 523 |  | 1 | 180 | - | $5.44 \times 10^{-8}$ |
|  | 3 | $10^{\text {b }}$ | 180 | 0.00-0.74 | - |
|  | 1 | 1 | 265 | - | $1.58 \times 10^{-8}$ |
|  | , | 3 | 265 | 1.78 | - |
|  | 1 | 0 | None | - | - |

${ }^{a}$ Calculated using CERVUS when there was more than one nonexcluded male and is the range over all offspring for the assigned male. Critical delta values for the relaxed and strict confidence levels are 1.44 and 3.10 , respectively.
${ }^{b}$ This is the total number of unique fathers over all of these offspring.
clutches $\left(t_{\mathrm{df}=5}=-1.641, P=.162\right.$; Table 1). A negative relationship exists between hatching success and multiple paternity ( $t_{\mathrm{df}=5}=2.254, P=.074$ ).

Some morphological differences were noted but were limited to differences among females of single- and multi-ple-sired clutches. Data on mass and CL were normally distributed and homoscedastic. Males who were assigned as fathers $(N=8)$ had larger CL than unassigned males ( $N=9$; $t_{\mathrm{df}=15}=2.400, P=.030$ ) but did not differ in mass $\left(t_{\mathrm{df}=15}=1.335, P=.202\right)$. Mass $\left(t_{\mathrm{df}=6}=-0.967, P=.371\right)$ and CL $\left(t_{\mathrm{df}=6}=-0.053, P=.960\right)$ were not significantly different between fathers of single-sire $(N=5)$ and multi-ple-sire clutches $(N=4)$. Mass $\left(t_{\mathrm{df}=5}=6.409, P=.001\right)$ and CL $\left(t_{\mathrm{df}=5}=2.682, P=.044\right)$ did differ between females inseminated by a single male $(N=5)$ and females inseminated by more than one male $(N=2)$, however. Females inseminated by multiple males on average weighed 1320 g less and had CLs 23.1 mm shorter than females inseminated by a single male (Table 1).

## Discussion

All mothers were genotyped, and no mismatches between known mother-offspring pairs were observed. The mother's contribution to each clutch was identified and removed, and all remaining alleles were designated as paternal alleles. Multiple paternity was observed in two of the seven clutches. In general, the percentage of clutches in which multiple paternity is found varies with species, and sometimes even among populations within a species. For those turtle species that have been studied, the percentage of multiply fertilized clutches ranges from $4 \%$ to $100 \%$ (Hoekert et al. 2002; Moore and Ball 2002; Pearse and Avise 2001). In this study,
$28.6 \%$ showed multiple paternity. This number may be conservative, however, as the ability of the loci used in this study to detect multiple paternity was only $87.6 \%$. Thus, for each clutch in which one father was detected, there was a $12.4 \%$ chance that another father went undetected. A full characterization of the frequency of multiple paternity in gopher tortoises will require a much more extensive survey including several populations from throughout the range and more individuals per population.

One offspring, in multiple sire clutch 529 , could not be assigned to any of the genotyped males. One possibility is that this individual was indeed the offspring of one of the surveyed males but appears different from either the mother or father as a result of new mutation. Because there are two loci each with a novel allele, mutation would have had to have happened independently at each of them concurrently, which is unlikely. A second, more reasonable, possibility is that three different males, one of which was not among the genotyped males, inseminated the female. Between 1988 and 1993, 42 adult males have been identified in the area, but we genotyped only 17 ( $41 \%$ ). A third possibility is that the female may be a recent immigrant into the area. Although all individuals in this area have been routinely marked (Mushinsky et al. 1994), the mother of the unassignable offspring was not. It is possible then that in addition to the sperm from the two local males the female was carrying sperm from at least one male outside the ERA. Given the possibility of long-term sperm storage in gopher tortoises, this latter scenario is possible. Regardless, multiple paternity is clearly observed in gopher tortoises and may constitute a frequent mating outcome. We cannot, however, differentiate between intraseasonal and temporal polyandry.

Reports of aggressive interactions between male gopher tortoises have provoked the question of whether or not these
duels influence a male's access to females. If winning males do receive greater access to females, then the victor's fitness would likely be higher. If the chances of winning are positively correlated with size, then larger males would inseminate more females and fertilize more eggs. In this study, one large male (43) fertilized two of the seven gravid females sampled (nearly $30 \%$ ) and 12 of the 53 eggs ( $23 \%$ ). The remaining eight fathers each fertilized only one female and from $1.2 \%$ to $20.7 \%$ of the eggs. Males that were assigned as sires of the clutches were significantly larger than the unassigned males, suggesting that larger males have a reproductive advantage. Larger males often have been reported as the winners of aggressive interactions (Douglass 1976). Here, we also observed aggressive male-male interactions during the breeding season near the entrances to female burrows. Thus, it is likely that the greater proportion of fertilization attributable to larger males is at least partially a result of winning aggressive encounters and gaining greater access to females.

Adult size also may be a significant factor regulating multiple paternity. Female, but not male, size was significantly different between single- and multiple-sired clutches. Females with multiple-sired clutches were lighter and shorter than females with single-sired clutches (Table 1). At least two explanations exist for this difference. The first involves male guarding behavior. Male-male duels and male-female courtships in gopher tortoises can take a considerable amount of time (Douglass 1976; McRae et al. 1981). Even males that are generally successful in aggressive encounters, therefore, may be temporally limited in the number of females with which they can mate. If males are limited to only a few females, then males may prefer, and preferentially guard, larger females that can produce more eggs (Pearse et al. 2002). If larger males are more likely to select and successfully guard large females, then large males should on average sire clutches from large females. The correlation ( $R=.852, P=.067$ ) of female versus male parent mass for single-sired clutches is consistent with this expectation (data available on request). Larger males preferentially guarding larger females also would result in smaller females either being unguarded or ineffectively guarded by small males. It seems possible then that this could result in smaller females being more frequently inseminated by multiple males. After having copulated with a female, a small male could be displaced by a larger male, which, depending on the female's ability to fend off the male, may result in a multiple-sired clutch. Although our sample size is somewhat small, our results are consistent with this in that the two females producing multiple-sired clutches were lighter and had shorter CL than all females with single-sired clutches (Table 1).

A second explanation for differences in female size for single- and multiple-sired clutches involves female receptiveness. In the field, females have been observed blocking their burrow entrances and rebuffing males performing courting behaviors (Douglass 1976; personal observation). Larger females may be better able to turn away attempts by courting males than are smaller females, especially when the courting males are large. If this were the case, then, again, we would expect the incidences of multiple paternity to be higher in
small females, as is observed. It is important to note that we cannot differentiate between explanations involving size versus age (i.e., experience) because age and size are strongly correlated. Smaller and lighter females are likely also to represent younger and less experienced females. Novice females may be engaging in different behaviors simply because they are inexperienced, and any difference may reflect experience and not size, per se. Younger females may also be behaviorally as well as physically less selective in choosing mates.

Regardless of the proximate cause of multiple-sired clutches, a clear result is that promiscuity, both in the form of polyandry and polygyny, is occurring in gopher tortoises. From a conservation standpoint, this is the most positive situation. Increases in the variance of reproductive success (i.e., polygyny) reduce the genetically effective population size $\left(N_{\mathrm{e}}\right)$ relative to the census size (Wright 1938). Reductions in $N_{\mathrm{e}}$ can lead to inbreeding and higher probabilities of expressing deleterious recessive alleles resulting in reduced fitness. Even so, the addition of polyandry and a long generation time (i.e., overlapping generations) likely are ameliorating factors (Nunney 1993) partially, if not completely, compensating for any reductions in $N_{\mathrm{e}}$. With these initial data, however, we cannot fully determine the evolutionary or conservation implications or how typical the ERA population is of other tortoise populations. However, it does appear that the reproductive output and probability of mating with more than one male are related to female size or experience (at least to the extent that they are correlated). Further studies using other populations and more individuals per population are necessary to fully characterize the consequences of the promiscuous mating system in gopher tortoises.

## Acknowledgments

This project was supported by a J. Landers Student Research Award to J.C.M. and Arcadia Wildlife Preserve, Inc. and National Science Foundation Grant in Systematics DEB 98-06905 to S.A.K. We thank Tonia Schwartz for her aid with laboratory procedures and three anonymous reviewers for helpful comments on an earlier draft. Animals were trapped and collected under Florida Fish and Wildlife Service permit WV01274, and tissue collection and handling procedures were approved by The University of South Florida IACUC permit number 1742.

## References

Auffenberg W, 1966. On the courtship of Gopherus polyphemus. Herpetologica 22:113-117.
Auffenberg W and Franz R, 1982. The status and distribution of the gopher tortoise (Gopherus polyphemus). In: North American tortoises: conservation and ecology (Bury RB, ed). Washington, DC: U.S. Department of Interior, Fish and Wildlife Service, Wildlife Research Report 12; 95-126.
Boglioli MD, Guyer C, and Michener WK, 2003. Mating opportunities of females gopher tortoises, Gopherus polyphemus, in relation to spatial isolation of females and their burrows. Copeia 2003:846-850.
Bureau of Economic and Business Research, 2000. Florida statistical Abstract. Gainesville, FL: University Presses of Florida.
Burke RL, Ewert MA, McLemore JB, and Jackson DR, 1996. Temperaturedependent sex determination and hatching success in the gopher tortoise (Gopherus polyphemus). Chelonian Conserv Biol 2:86-88.

Butler JA and Hull TW, 1996. Reproduction of the tortoise, Gopherus polyphemus, in northeastern Florida. J Herpetol 30:14-18.
Byrne PG and Roberts JD, 2000. Does multiple paternity improve the fitness of the frog Crinia georgiana? Evolution 54:968-973.
Crim JL, Spotila D, Spotila R, O’Connor M, Reina R, and Williams CJ, 2002. The leatherback turtle Dermochelys coriacea, exhibits both polyandry and polygyny. Mol Ecol 11:2097-2106.
Dallas JF, 1992. Estimation of microsatellite mutation rates in recombinant inbred strains of mouse. Mamm Genome 5:32-38.
Demuth JP, 2001. The effects of constant and fluctuating incubation temperature on sex determination, growth, and performance in the tortoise Gopherus polyphemus. Can J Zool 79:1609-1620.
Diemer JE, 1986. The ecology and management of the gopher tortoise in the southeastern United States. Herpetologica 42:125-133.
Diemer JE and Moore CT, 1994. Reproductive biology of gopher tortoises in north-central Florida. In: Biology of North American tortoises (Bury RB and Germano DJ, eds). Washington, DC: U.S. Fish and Wildlife Service, Fish and Wildlife Research Report 13; 129-137.
Douglass JF, 1976. The mating system of the gopher tortoise, Gopherus polyphemus, in southern Florida (MS thesis). Tampa, FL: University of South Florida.
Dudash M and Fenster CB, 2000. Inbreeding and outbreeding depression in fragmented populations. In: Genetics, demography and viability of fragmented populations (Young AG and Clarke GM, eds). New York: Cambridge University Press; 35-53.
Epperson DM and Heise CD, 2003. Nesting and hatchling ecology of gopher tortoises (Gopherus polyphemus) in southern Mississippi. J Herpetol 37:315324.

Ewert MA and Legler JM, 1978. Hormonal induction of oviposition in turtles. Herpetologica 34:314-318.
Ewing HE, 1943. Continued fertility in female box turtles following mating. Copeia 1943:112-114.
Florida Fish and Wildlife Conservation Commission, 2005. Florida's endangered species, threatened species, and species of special concern. Available at: http://www.wildflorida.org. Accessed April 27, 2005.
Gilpin ME and Soulé ME, 1986. Minimum viable populations; processes of species extinction. In: Conservation biology (Soulé ME, ed). Sunderland, MA: Sinauer Associates, Inc.; 19-34.
Hailman JP, 1991. Notes on the aggressive behavior of the gopher tortoise. Herpetol Rev 22:87-88.
Hanotte O, Burke T, Armour JAL, and Jeffreys AJ, 1991. Hypervariable minisatellite DNA sequences in the Indian peafowl Pavo cristatus. Genomics 9:587-597.
Herrmann BG and Frischauf A, 1987. Isolation of genomic DNA. Methods Enzymol 152:180-182.
Hoekert WEJ, Neuféglise H, Schouten AD, and Menken SBJ, 2002. Multiple paternity and female-biased mutation at a microsatellite locus in the olive ridley sea turtle (Lepidochelys olivacea). Heredity 89:107-113.
Inskipp T and Gillett HJ, 2003. Checklist of CITES species. Compiled by UNEP-WCMC, CITES Secretariat, Geneva, Switzerland and UNEPWCMC, Cambridge, UK; 339 pp . and CD-ROM.
Iverson JB, 1980. The reproductive biology of Gopherus polyphemus (Chelonia: Testudinidae). Am Midl Nat 103:353-359.
Jessop TS, FitzSimmons NN, Limpus CJ, and Whittier JM, 1999. Interactions between behavior and plasma steroids within the scramble mating system of the promiscuous green turtle, Chelonia mydas. Horm Behav 36: 86-97.
Jones AG, Rosenqvist G, Berglund A, and Avise JC, 1999. The genetic mating system of a sex-role-reversed pipefish (Syngnathus typble): a molecular inquiry. Behav Ecol Sociobiol 46:357-365.

Kautz RS, 1993. Trends in Florida wildlife habitat 1939-1987. Fla Sci 56: 7-24.
Kempenaers B, Verheyen GR, Vandenbroeck M, Burke T, Vanbroeckhoven C, and Dhondt AA, 1992. Extra-pair paternity results from female preference for high-quality males in the blue tit. Nature 357:494-496.
Lande R and Barrowclough GF, 1987. Effective population size, genetic variation, and their use in population management. In: Viable populations for conservation (Soulé ME, ed). Cambridge, MA: Cambridge University Press; 87-123.
Madsen T, Shine R, Loman J, and Hakansson T, 1992. Why do female adders copulate so frequently? Nature 355:440-441.
Marshall TC, Slate J, Kruuk LEB, and Pemberton JM, 1998. Statistical confidence for likelihood-based paternity inference in natural populations. Mol Ecol 7:639-655.
McRae WA, Landers JL, and Garner JA, 1981. Movement patterns and home range of the gopher tortoise. Am Midl Nat 106:165-179.
Moore MK and Ball RM Jr, 2002. Multiple paternity in loggerhead turtle (Caretta caretta) nests on Melbourne Beach, Florida: a microsatellite analysis. Mol Ecol 11:281-288.
Mushinsky HR, 1985. Fire and the Florida sandhill herpetofaunal community: with special attention to responses of Cnemidophorus sexlineatus. Herpetologica 41:333-342.
Mushinsky HR, 1992. Natural history and abundance of southeastern fivelined skinks, Eumeces inexpectatus, on a periodically burned sandhill in Florida. Herpetologica 48:307-312.
Mushinsky HR and McCoy ED, 1994. Comparison of gopher tortoise populations on islands and on the mainland in Florida. In: Biology of North American tortoises (Bury RB and Germano DJ, eds). Washington, DC: U.S. Fish and Wildlife Service, Fish and Wildlife Research Report 13: 39-47.
Mushinsky HR, Wilson DS, and McCoy ED, 1994. Growth and sexual dimorphism of Gopherus polyphemus in central Florida. Herpetologica 50: 119-128.
Noss RF, 1989. Longleaf pine and wiregrass: keystone components of an endangered ecosystem. Nat Areas J 9:211-213.
Nunney L, 1993. The influence of mating system and overlapping generations on effective population size. Evolution 47:1329-1341.
Orsetti DM and Rutowski RL, 2003. No material benefits, and a fertilization cost, for multiple mating by female leaf beetles. Anim Behav 66:477-484.
Otter K and Ratcliffe L, 1996. Female initiated divorce in a monogamous songbird: abandoning mates for males of higher quality. Proc R Soc Lond B 263:351-354.
Palmer KS, Rostal DC, Grumbles JS, and Mulvey M, 1998. Long-term sperm storage in the desert tortoise (Gopherus agassizii). Copeia 1998:702-705.
Pearse DE and Avise JC, 2001. Turtle mating systems: behavior, sperm storage, and genetic paternity. J Hered 92:206-211.
Pearse DE, Janzen FJ, and Avise JC, 2002. Multiple paternity, sperm storage, and reproductive success of female and male painted turtles (Chrysemys picta) in nature. Behav Ecol Sociobiol 51:164-171.
Queller DC, Strassmann JE, and Hughes CR, 1993. Microsatellites and kinship. Trends Ecol Evol 8:285-288.
Roques S, Diaz-Paniagua C, and Andreu AC, 2004. Microsatellite markers reveal multiple paternity and sperm storage in the Mediterranean spurthighed tortoise, Testudo graeca. Can J Zool 82:153-159.
Schwartz TS, Osentoski M, Lamb T, and Karl SA, 2003. Microsatellite loci for the North American tortoises (genus Gopherus) and their applicability to other turtle species. Mol Ecol Primer 3:283-286.
Strausberger BM and Ashley MV, 2003. Breeding biology of brood parasitic brown-headed cowbirds (Molothrus ater) characterized by parent-offspring and sibling-group reconstruction. Auk 120:433-445.
U.S. Fish and Wildlife Service, 1986. Endangered and threatened wildlife and plants: determination of threatened status for the gopher tortoise (Gopherus polyphemus). Fed Regist 52:25376-25380.
Valenzuela N, 2000. Multiple paternity in side-neck turtle Podocnemis expansa: evidence from microsatellite DNA data. Mol Biol 9:99-105.
Watson PJ, 1998. Multi-male mating and female choice increase offspring growth in the spider Neriene litigiosa (Linymphiidae). Anim Behav 55:387-403.
Westneat DF, Fredrick PC, and Wiley RH, 1987. The use of genetic markers to estimate the frequency of successful alternative reproductive tactics. Behav Ecol Sociobiol 21:35-45.
Wilson DS, Mushinsky HR, and McCoy ED, 1991. Relationship between gopher tortoise body size and burrow width. Herpetol Rev 22:122-124.

Wright JS, 1982. The distribution and population biology of the gopher tortoise (Gopherus polyphemus) in South Carolina (MS thesis). Clemson, SC: Clemson University.
Wright S, 1938. Size of population and breeding structure in relation to evolution. Science 87:430-431.

Zeh JA and Zeh DW, 1996. The evolution of polyandry I: intragenomic conflict and genetic incompatibility. Proc R Soc Lond B 263:1711-1717.

Received July 22, 2005
Accepted November 30, 2005
Corresponding Editor: Brian Bowen

